

Curso Avanzado de Actualización en Onco-hematología por
Citometría de flujo,
Buenos Aires, 30 de mayo de 2011

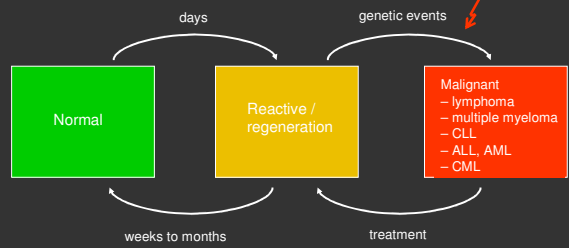
Proyecto EuroFlow: objetivos, resultados y perspectivas futuras

Un nuevo concepto en diagnóstico or citometría de flujo

Jacques J.M. van Dongen & Alberto Orfao
por el grupo



Discrimination between normal and malignant immune cells in blood, bone marrow and lymphoid tissues



Laboratory methods:

- Cytomorphology and immunophenotyping
- Molecular diagnostics, e.g. PCR-based clonality diagnostics and molecular classification via detection of oncogenetic defects

Diagnostics for hematological malignancies

1. Making the diagnosis

Normal ↔ reactive/regenerating ↔ malignant

Annually > 300,000 new patients with a hematological malignancy in developed countries

2. Classification of hematopoietic malignancies

- relation with prognosis
- relevance of risk-group definition in treatment protocols

→ Based on differentiation characteristics and particularly on chromosome aberrations, resulting in fusion gene transcripts or aberrantly (over) expressed genes

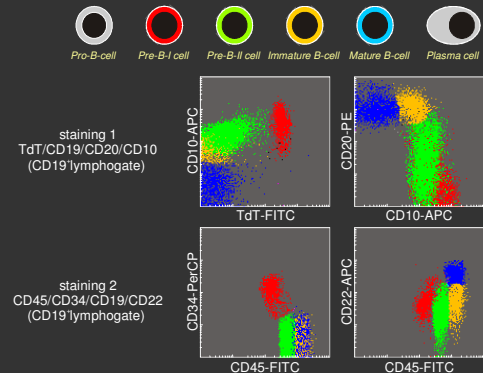
3. Evaluation of treatment effectiveness

Detection of minimal residual disease (MRD):

MRD-based risk-group stratification (treatment reduction or treatment intensification)

Annually > 400,000 follow-up samples in leukemia patients (ALL, AML, CML)

Identification of different B-cell subpopulations in childhood BM



E. G. van Lochem et al., *Cytometry Part B* 2004; 60B: 1-13.

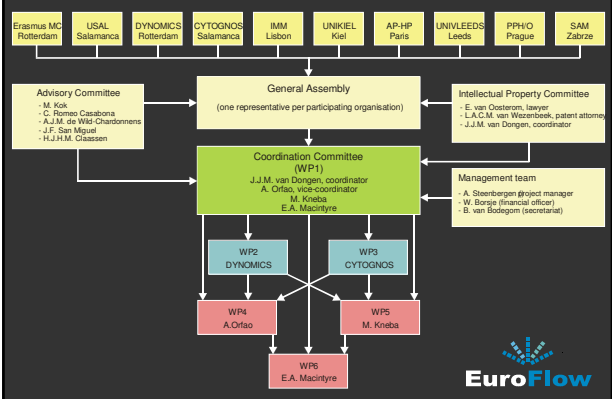
Flow cytometric immunophenotyping of normal and malignant leukocytes

Gaps and areas for improvement (Status in 2005)

- no technical standardization in flow cytometry
- no guidelines for selection of the appropriate antibody clones
- virtually no new markers introduced over a decade (stand-still in development)
- many oncoproteins (including fusion proteins) not yet included in immunostaining protocols;
- 3- and 4-color flow cytometry has many limitations: limited sensitivity and limited specificity
- management of large data files from multiple samples is complex and time-consuming; new software needed for:
 - fast and easy analysis of data;
 - automated patient reports;
 - introduction of flow data into electronic hospital systems.

→ International collaboration between academia and industry

Management structure of EuroFlow Consortium



Achievements of the EuroFlow Consortium

- Multicolor flow cytometry (≥8 colors) with full technical standardization
- inclusion of violet laser and selection of appropriate fluorochromes
 - standardization of instrument settings and laboratory protocols
 - detailed testing and comparison of antibody clones and conjugated antibodies (multiple companies)
- Implementation and development of novel software
- fast and easy handling of large data files (including automated pattern recognition)
 - combining multiple tubes: calculation and APS view
 - mapping of diagnosis and follow-up leukemia samples against templates of "normal/control" samples
- Development of 8-color antibody protocols for diagnosis, classification and monitoring of hematological malignancies
- screening tubes (include recognition of normal leukocyte subsets)
 - multi-tube panels for diagnosis and classification per disease category
 - special tubes for MRD monitoring per disease category

Standardization in diagnostic flow cytometry

- Standardization according to literature generally refers to:
- lists of CD codes and markers per disease category
 - rarely a specific antibody is recommended and (almost) never a fluorochrome is proposed

HOWEVER: Standardization according to GLP guidelines demands for much higher levels of standardization

- EuroFlow standardization aims at:
- usage of comparable flow cytometers (3 lasers and ≥ 8 colors)
 - full standardization of instrument settings (e.g. based on standard beads)
 - standardized laboratory protocols and immunostaining procedures (SOP's)
 - careful selection of optimal antibody clones per marker/CD code
 - selection of optimal 8-color antibody combinations and fluorochromes
 - design of combinations of multiple 8-color tubes: estimation and APS view
 - new software for fast and easy data analysis with automated pattern recognition
 - recognition of normal and abnormal leukocyte subsets (complete differentiation pathways) with the same immunostaining protocols
 - mapping of new patient samples against large data base of earlier collected patient samples, analyzed with the same immunostaining protocol

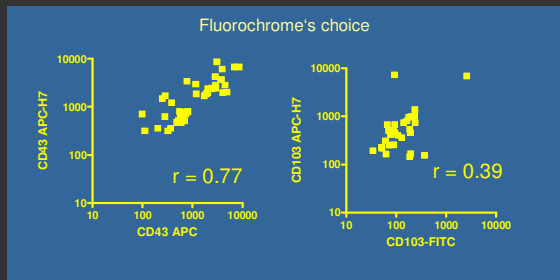
Fluorochromes for 8-color flow cytometric immunophenotyping

Fluorochrome	Excitation Peak (nm)	Emission Peak (nm)	Lasers		
			Violet	Argon	Helium-Neon
Pacific Blue	405	455	+		
AmCyan	405	490	+		
Pacific Orange	405	550	+		
Marina Blue	365	460		+	
FITC	495	520		+	
Phycocerythrin (PE)	565	575		+	
PE Texas Red	565	615		+	
PerCP	488	678		+	
PerCP-Cy5.5	488	695		+	
PE-Cy7	565	770		+	
Allophycocyanin (APC)650		660			+
Alexa 700	635	720			+
APC-H7	650	770			+

Fluorochromes for 8-color flow cytometric immunophenotyping

Fluorochrome	Excitation Peak (nm)	Emission Peak (nm)	Lasers		
			Violet	Argon	Helium-Neon
Pacific Blue/Horizon	405	455	+		
AmCyan	405	490	+		
Pacific Orange/Horizon	405	550	+		
Marina Blue	365	460		+	
FITC	495	520		+	
Phycocerythrin (PE)	565	575		+	
PE Texas Red	565	615		+	
PerCP	488	678		+	
PerCP-Cy5.5	488	695		+	
PE-Cy7	565	770		+	
Allophycocyanin (APC)650		660			+
Alexa 700	635	720			+
APC-H7	650	770			+

Construction of EuroFlow panels Fluorochrome conjugates, antibody panels, and antibody combinations



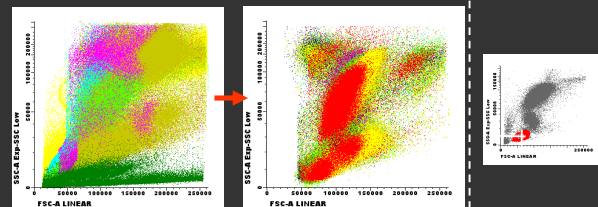
Responsible scientist: Sebastian Bottcher



Synchronized light scatter experiments

"Local" settings

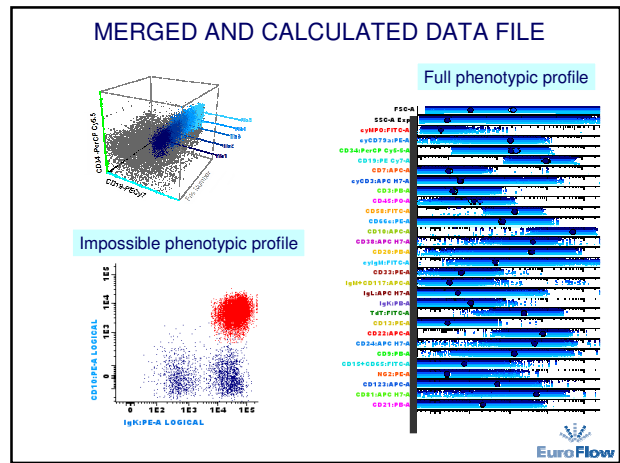
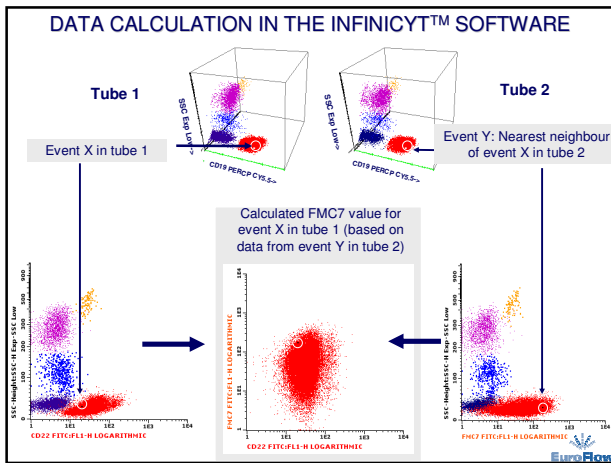
EuroFlow settings



7 different normal PB samples acquired in 7 different centers

Normal PB samples processed according to the standardized EuroFlow sample preparation protocol





MERGED & CALCULATED LISTMODE DATA FILE

Parameters/File	Files					
	1	2	3	4	5	6
FSC-A	C	C	C	C	C	C
SSC-A	C	C	C	C	C	C
KAPPA:FITC-A	R	E	E	E	E	E
LAI	R	R	R	R	R	R
CD	C	C	C	C	C	C
Dg	R	E	E	E	E	E
CD	C	C	C	C	C	C
CD	C	C	C	C	C	C
CD103:PerCP-Cy5.5	E	R	E	E	E	E
CD103:FITC-A	E	R	E	E	E	E
CD10:PE-A	E	R	E	E	E	E
CD43:APC-A	E	R	E	E	E	E
CD81:FITC-A	E	R	E	E	E	E
CD75b:PE-A	E	R	E	E	E	E
CD2	E	R	E	E	E	E
CD3	E	R	E	E	E	E
CD4	E	R	E	E	E	E
CD8	E	R	E	E	E	E
CD25:PE-A	E	R	E	E	E	E
CD138:PerCP-Cy5-5-A	E	R	E	E	E	E

6 Files x 10 parameters x 100,000 events

1 File x 25 parameters x 600,000 events

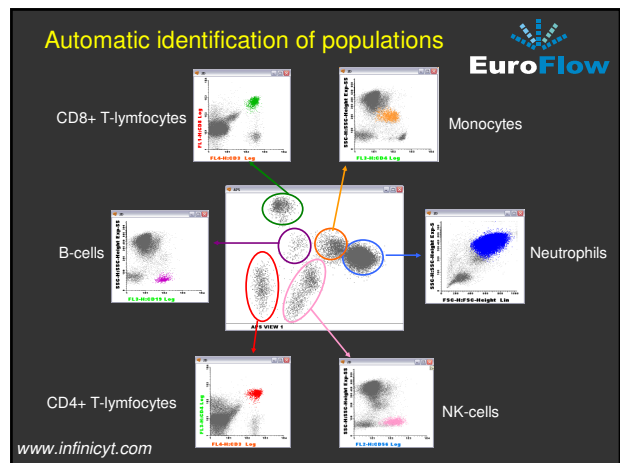
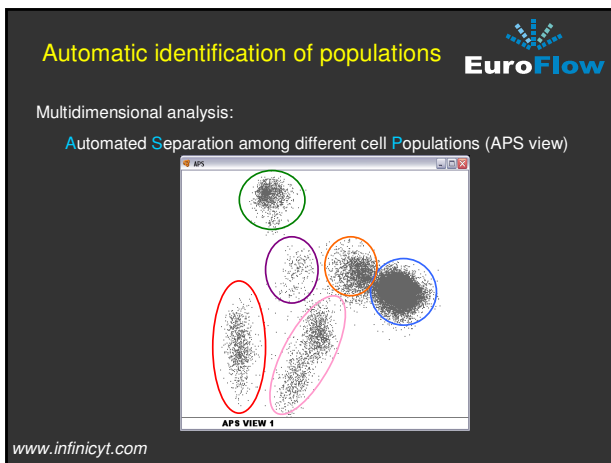
Parameter coding:
C = Common
R = Measured
E = Calculated

"Merging" by Infinicyt™ program

Integration of results from multiple tubes based on 4 to 6 common parameters per tube

Colors	No. of tubes	Total no. of antibodies	No. of common parameters antibodies	Scatter	End result no. of parameters
4-colors	8	32	2	2	20 (18+2)
6-colors	6	36	3	2	23 (21+2)
8-colors	4	32	4	2	22 (20+2)
8-colors	4	32	3	2	25 (23+2)

Development of 8-color multi-tube antibody protocols (3 or 4 antibodies in common per tube in each protocol)



APS Procedure for AUTOMATIC ANALYSIS

VISUALIZATION OPTIONS

Dots
Dots / Mean
Mean

APS Procedure for groups of patients

APS Dots View
APS Means View

Group of patients with the same panel/protocol applied and same disease category

APS Procedure for groups of patients

Each dot = patient/sample

Group of patients with same panel/protocol applied and 2 different disease categories

Results of synchronized experiments

APS view of 30 merged data files from different centers

CVMFI subset		
PacBlue channel	Lym - CD20pos	16,90%
PacOr channel	Lym - CD45 pos	15,52%
FTTC channel	Lym - CD3+ CD8pos	16,93%
PE channel	Lym - CD3+ CD27pos	27,95%
PerCPcy55 chann	Lym - CD3+ CD4pos	28,39%
PECy7 channel	Lym - CD19 pos	15,40%
APC channel	CD45pos - CD14 pos	22,68%
APC-H7 channel	Lym - CD3 pos	48,35%

EuroFlow

5th EuroFlow Educational Workshop,
Paris, FR, 9 March 2011

EuroFlow

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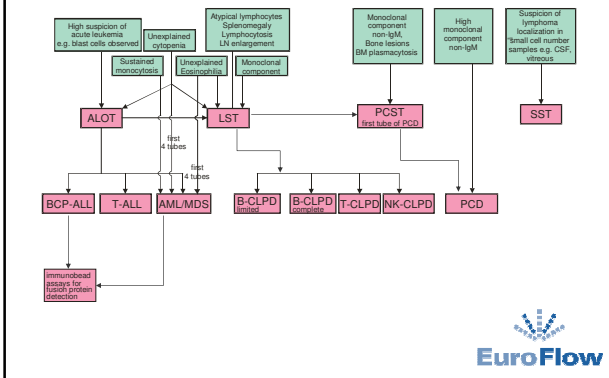
EuroFlow antibody protocols

EuroFlow

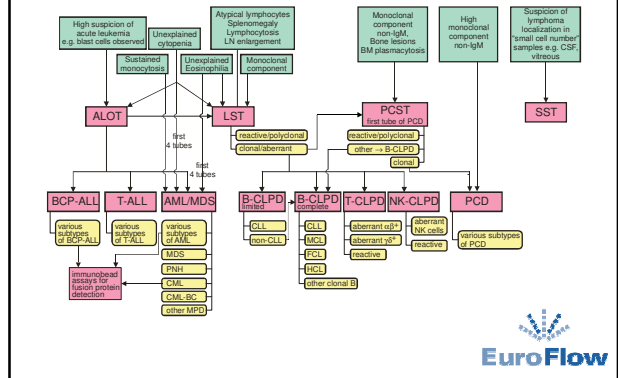
Development of 8-color multi-tube antibody protocols
(3 or 4 antibodies in common per tube in each protocol)

1. Screening tubes (include recognition of normal leukocyte subsets)
 - Acute leukemia orientation tube (ALOT): 1 tube (L Lhermitte)
 - Lymphoid screening tube (LST): 1 tube (J Flores Montero)
 - Small sample screening tube (SST): 1 tube (AW Langerak)
 - Plasma cell dyscrasia tubes (PCD): 2 tubes (J Flores Montero)
2. Multi-tube panels for characterization per disease category
 - B-cell precursor ALL (BCP-ALL) protocol: 4 tubes (L Lhermitte)
 - T-cell ALL (T-ALL) protocol: 4 tubes (V Asnafi)
 - AML/MDS protocol: 7 tubes (VHJ van der Velden)
 - B chronic lymphoproliferative diseases (B-CLPD): 5 tubes (S Böttcher)
 - T chronic lymphoproliferative diseases (T-CLPD): 6 tubes (J Almeida)
 - NK chronic lymphoproliferative diseases (NK-CLPD): 3 tubes (J Almeida)

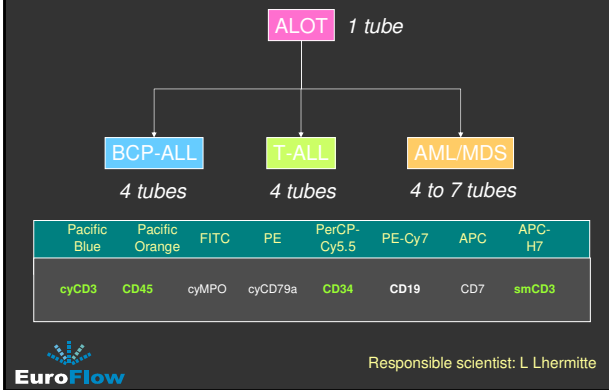
Algorithm for EuroFlow antibody panels in hemato-oncology



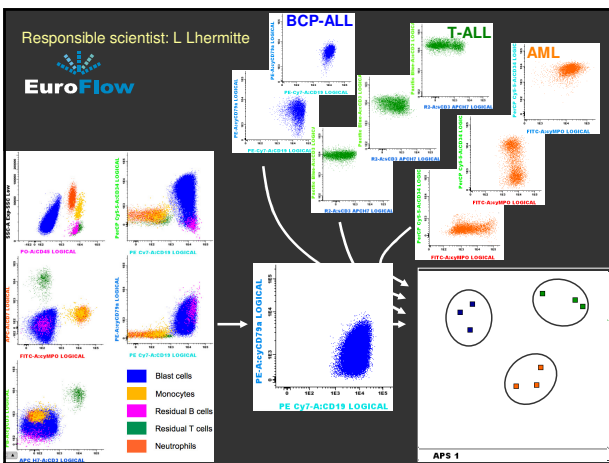
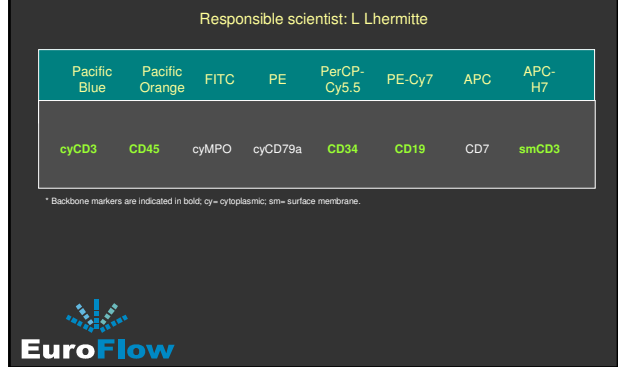
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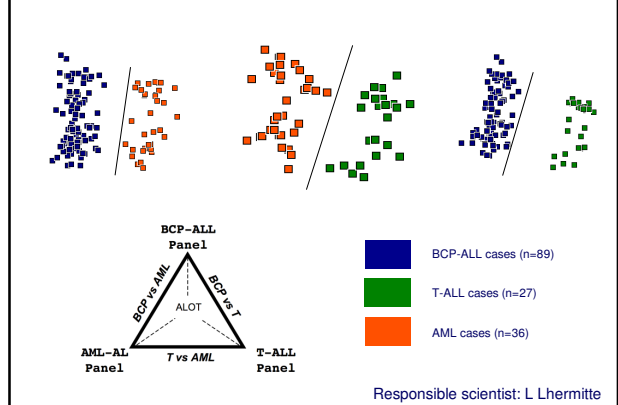
Acute leukemia orientation tube (ALOT)

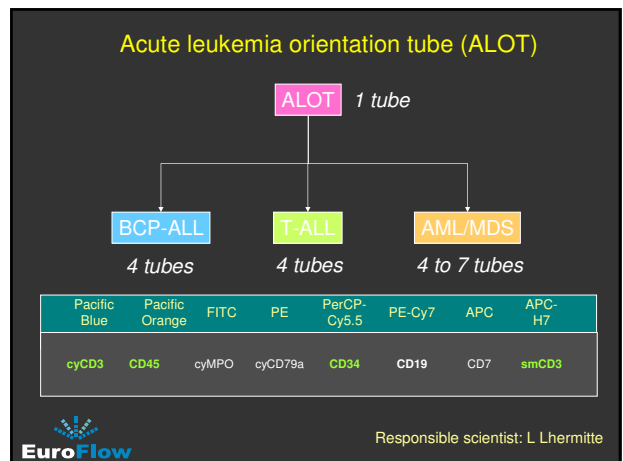
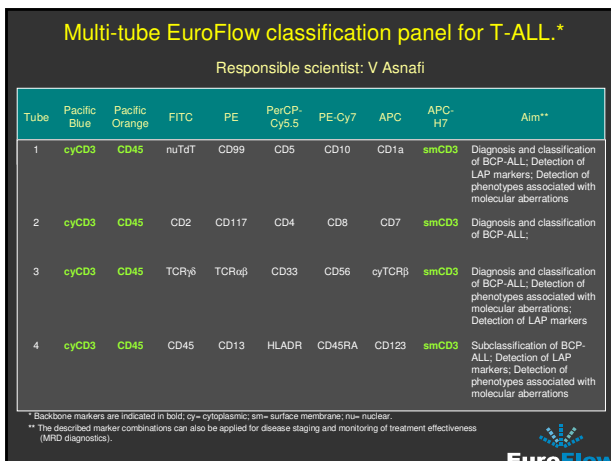
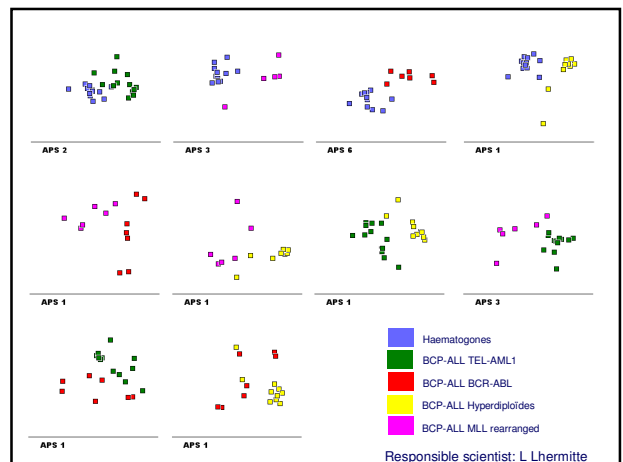
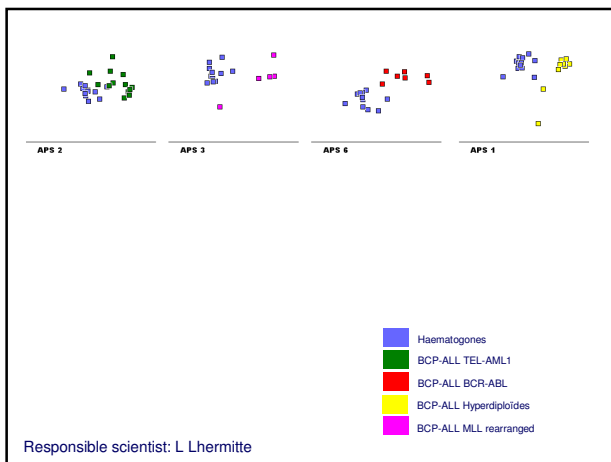
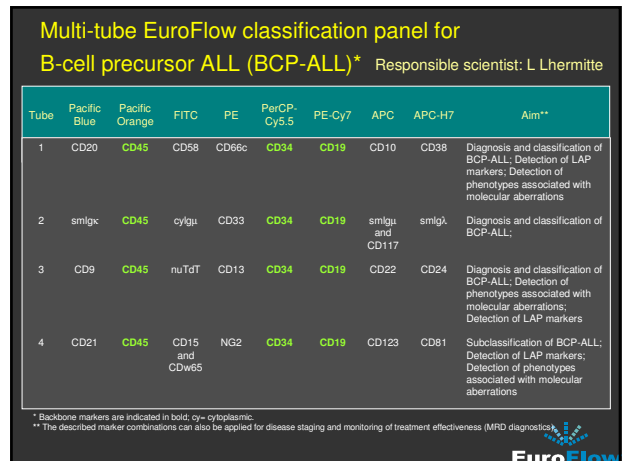
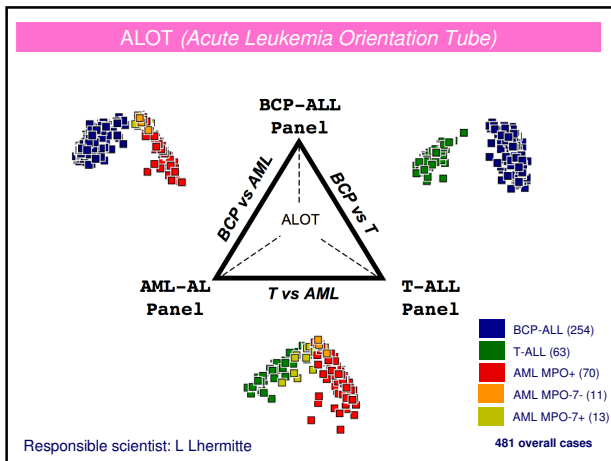


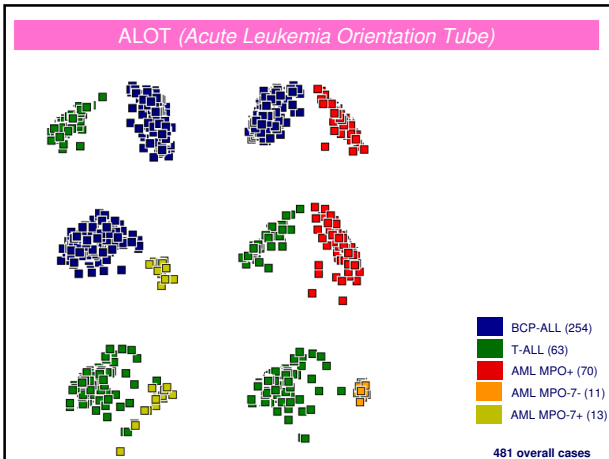
Single tube EuroFlow screening tube for acute leukemias Acute Leukemia Orientation Tube (ALOT)*



ALOT (Acute Leukemia Orientation Tube)





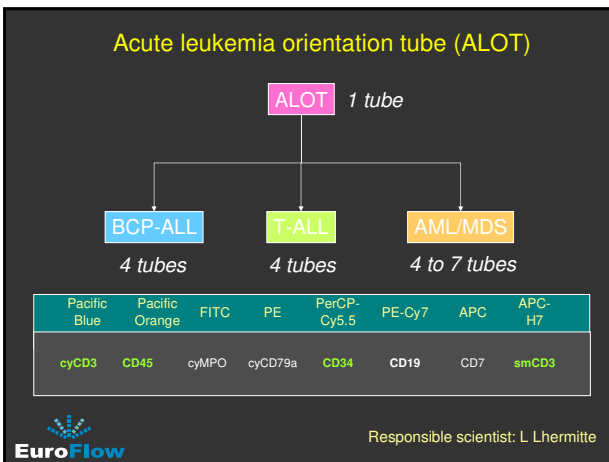


Multi-tube EuroFlow classification panel for B-cell precursor ALL (BCP-ALL)*

Responsible scientist: L Lhermitte

Tube	Pacific Blue	Pacific Orange	FITC	PE	PerCP-Cy5.5	PE-Cy7	APC	APC-H7	Aim**
1	CD20	CD45	CD58	CD66c	CD34	CD19	CD10	CD38	Diagnosis and classification of BCP-ALL; Detection of LAP markers; Detection of phenotypes associated with molecular aberrations
2	smIgκ	CD45	cyIgμ	CD33	CD34	CD19	smIgμ and CD117	smIgλ	Diagnosis and classification of BCP-ALL;
3	CD9	CD45	nuTdT	CD13	CD34	CD19	CD22	CD24	Diagnosis and classification of BCP-ALL; Detection of phenotypes associated with molecular aberrations; Detection of LAP markers
4	CD21	CD45	CD15 and CDw65	NG2	CD34	CD19	CD123	CD81	Subclassification of BCP-ALL; Detection of LAP markers; Detection of phenotypes associated with molecular aberrations

* Backbone markers are indicated in bold; cy- cytoplasmic.
 ** The described marker combinations can also be applied for disease staging and monitoring of treatment effectiveness (MRD diagnostics)



5th EuroFlow Educational Workshop, Paris, FR, 9 March 2011

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Multicolor flow cytometry (≥8 colors) with full technical standardization

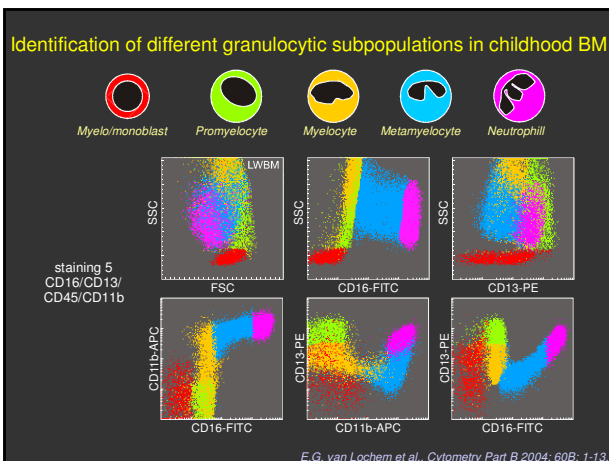
- inclusion of violet laser and selection of appropriate fluorochromes
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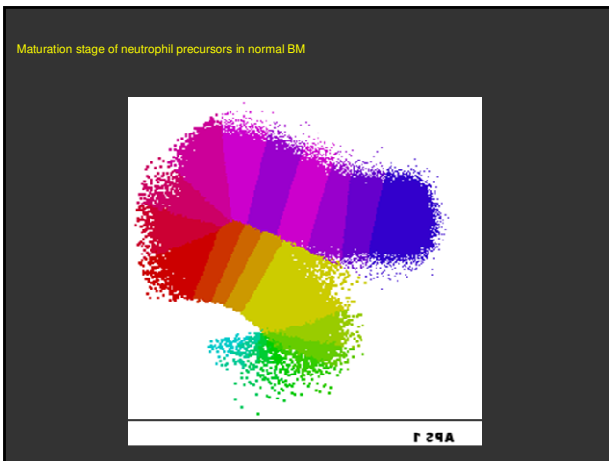
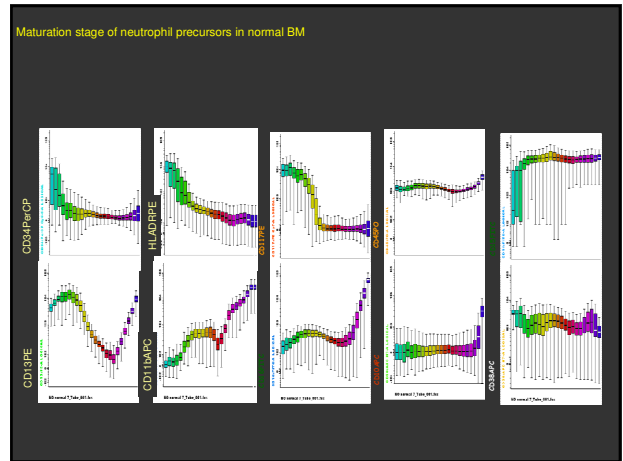
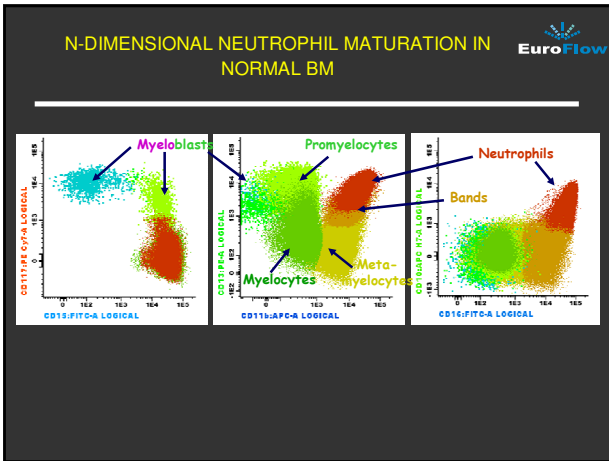
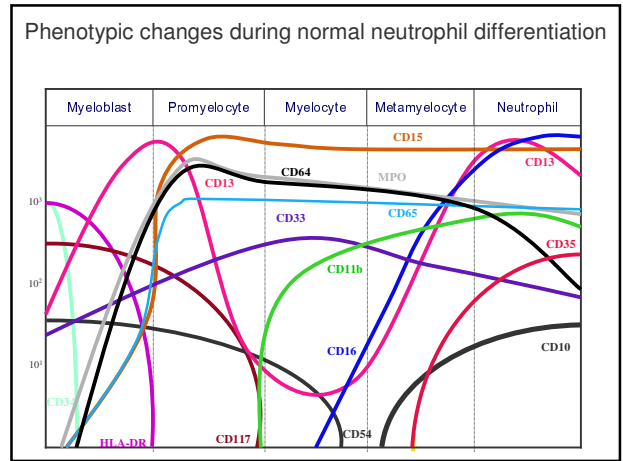
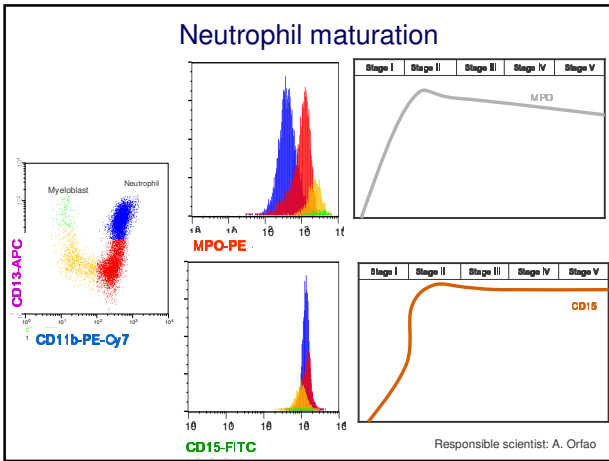


Multi-tube EuroFlow classification panel for AML/MDS

Responsible scientist: VHJ van der Velden

Tube	Pacific Blue	Pacific Orange	FITC	PE	PerCP-Cy5.5	PE-Cy7	APC	APC-H7	Aim**
1	HLADR	CD45	CD16	CD13	CD34	CD117	CD11b	CD10	Diagnosis and subclassification of AML and PNH especially focussed on neutrophilic lineage
2	HLADR	CD45	CD35	CD64	CD34	CD117	IREM2	CD14	Diagnosis and subclassification of AML and PNH especially focussed on monocytic lineage
3	HLADR	CD45	CD36	CD105	CD34	CD117	CD33	CD71	Diagnosis and subclassification of AML especially focussed on erythroid lineage
4	HLADR	CD45	nuTdT	CD56	CD34	CD117	CD7	CD19	Aberrant expression of lymphoid-associated markers and abnormal lymphoid maturation

* Further information about the markers and the availability of hybridoma clones is summarized in Appendix A. Backbone markers are indicated in bold; nu- nuclear.
 ** The described marker combinations might also be applied for disease staging and monitoring of treatment effectiveness (MRD diagnostics)



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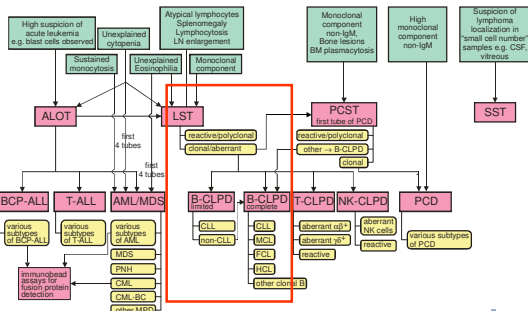
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Algorithm for EuroFlow antibody panels in hemato-oncology



Multi-tube EuroFlow classification combinations for B-cell chronic lymphoproliferative diseases (B-CLPD) *

Responsible scientist: S Böttcher supported by A Rawstron and J Flores Montero

Tube	Pacific Blue	Pacific Orange	FITC	PE	PerCP-Cy5.5	PE-Cy7	APC	APC-H7	Aim**
1	CD20 and CD4	CD45	CD8 and smlgλ	CD56 and smlgκ	CD5	CD19 and TCRγδ	smCD3 8	CD38	LST tube; detection of B-CLPD
2	CD20	CD45	CD23	CD10	CD79b	CD19	CD200	CD43	Identification of CLL vs other B-CLPD cases, when combined with LST

* Backbone markers are indicated in bold; sm= surface membrane.
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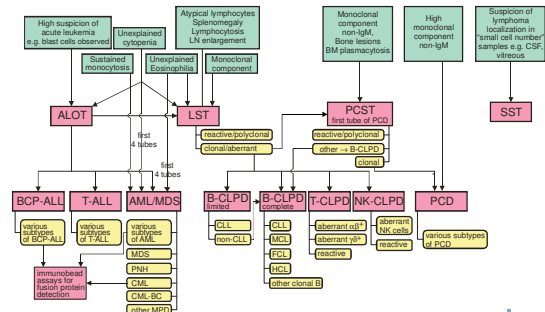
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Tube	Pacific Blue	Pacific Orange	FITC	PE	PerCP-Cy5.5	PE-Cy7	APC	APC-H7	Aim**
1	CD20 and CD4	CD45	CD8 and smlgλ	CD56 and smlgκ	CD5	CD19 and TCRγδ	smCD3 8	CD38	LST tube; detection of B-CLPD
2	CD20	CD45	CD23	CD10	CD79b	CD19	CD200	CD43	Identification of CLL vs other B-CLPD cases, when combined with LST
3	CD20	CD45	CD31	LAI1	CD11c	CD19	smlgμ	CD81	Further subclassification of non-CLL B-CLPD, e.g. FCL, MCL, FL, MZL, LPL, DLBCL and other B-CLPD
4	CD20	CD45	CD103	CD95	CD22	CD19	CXCR5	CD49d	
5	CD20	CD45	CD62L	CD39	HLADR	CD19	CD27		

* Backbone markers are indicated in bold; sm= surface membrane.
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Algorithm for EuroFlow antibody panels in hemato-oncology



Technical aspects of EuroFlow protocols: instrument settings, fluorochrome choice, standardization

T. Kalina¹, J. Flores-Montero², Q. Lécresse², M. Cullen³, L. Lhermitte⁴, L. Sedek⁵, A. Mendonça⁶, S. Böttcher⁷, J. te Marvalde⁸, Mejstříková, O. Hrušák¹, J.J.M. van Dongen⁹, and A. Orfao²

On behalf of the EuroFlow Consortium (EU-FP6, LSHB-CT-2006-018708)

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4. Department of Hematology, Hôpital Necker, Paris, FR
5. Department of Pediatric Hematology and Oncology, Medical University of Silesia, Zabrze, PL;
6. Department of Hematology, Instituto Português de Oncologia, Lisbon, PT;
7. 2nd Department of Medicine, University Klinik Schleswig-Holstein, Kiel, DE;
8. Department of Immunology, Erasmus MC, Rotterdam, NL;

To be published in: *Leukemia* 2011; 25: xxxx-xxxx



EuroFlow antibody panels for standardized n-dimensional flow cytometric immunophenotyping of normal, reactive and malignant leukocytes

J.J.M. van Dongen, L. Lhermitte, S. Böttcher, J. Almeida, V.H.J. van der Velden, J. Flores-Montero, A. Rawstron, V. Asnafi, Q. Lécresse, P. Lucio, E. Mejstříková, T. Szczepański, T. Kalina, R. de Tute, M. Brüggemann, L. Sedek, M. Cullen, A.W. Langerak, A. Mendonça, E. Macintyre, M. Martin-Ayuso, O. Hrusak, M.B. Vidriales, and A. Orfao

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- standardization of instrument settings and laboratory protocols
- detailed testing and comparison of antibody clones and conjugated antibodies (multiple companies)

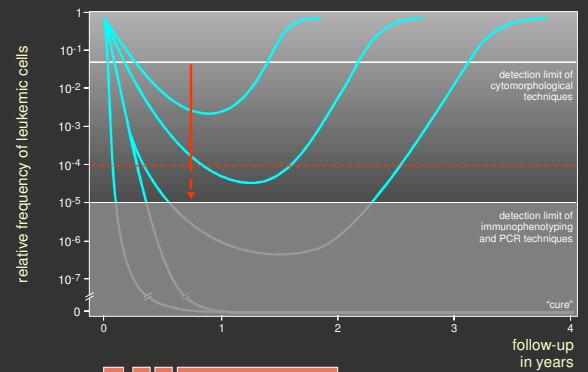
Implementation and development of novel software

- fast and easy handling of large data files (including automated pattern recognition)
- combining multiple tubes: calculation and APS view
- mapping of diagnosis and follow-up leukemia samples against templates of "normal/control" samples

Development of 8-color antibody protocols for diagnosis, classification and monitoring of hematological malignancies

- screening tubes (include recognition of normal leukocyte subsets)
- multi-tube panels for diagnosis and classification per disease category
- special tubes for MRD monitoring per disease category

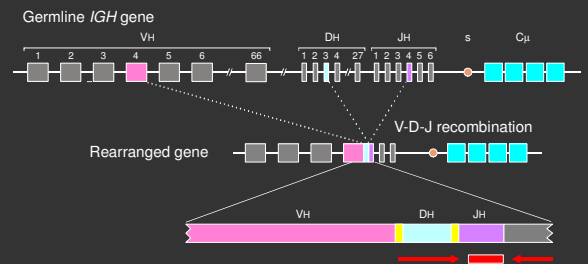
Detection of minimal residual disease (MRD)



Detection of minimal residual disease in ALL

Technique	Applicability	Detection limit	Remark
Flow cytometry (4 colors)	BCP-ALL: 85% T-ALL: 90%	$(10^{-3}) - 10^{-4}$	Fast, but variable sensitivity because of similarities between normal (regenerating) cells and malignant cells
PCR of Ig/TCR genes	BCP-ALL: 95% T-ALL: 95%	$10^{-4} - 10^{-5}$	Time consuming and relatively expensive (junctional region sequencing), but applicable in $\geq 95\%$ of lymphoid malignancies
PCR of fusion transcripts	BCP-ALL: 40% T-ALL: 25%	$10^{-4} - 10^{-6}$	Limited applicability in ALL, but potentially useful in specific subgroups, e.g. BCR-ABL cases in specific protocols

PCR analysis of Ig/TCR genes



EuroMRD: European Study Group on MRD detection (ESG-MRD-ALL)



AIMS of EuroMRD (focus on PCR analysis of Ig/TCR genes):

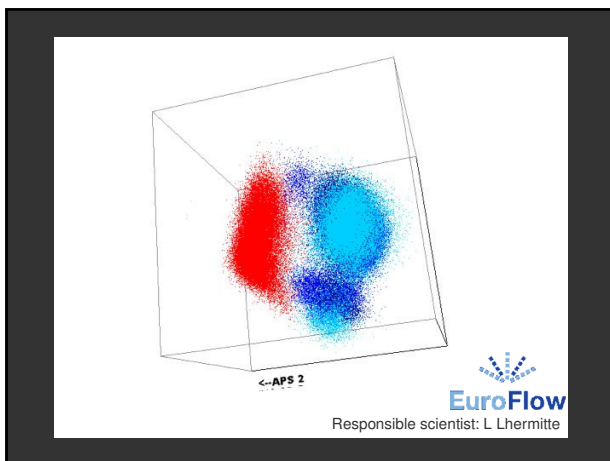
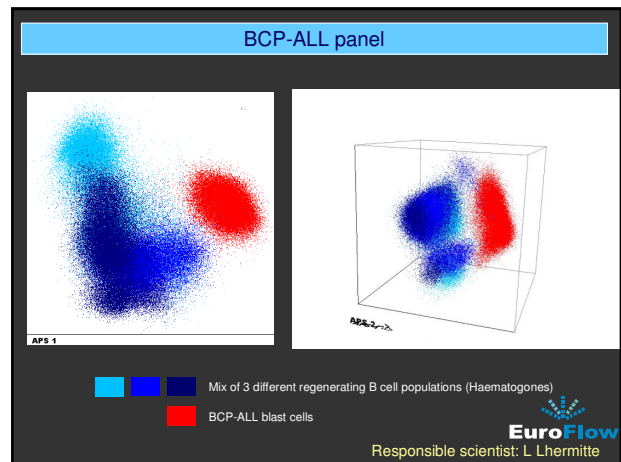
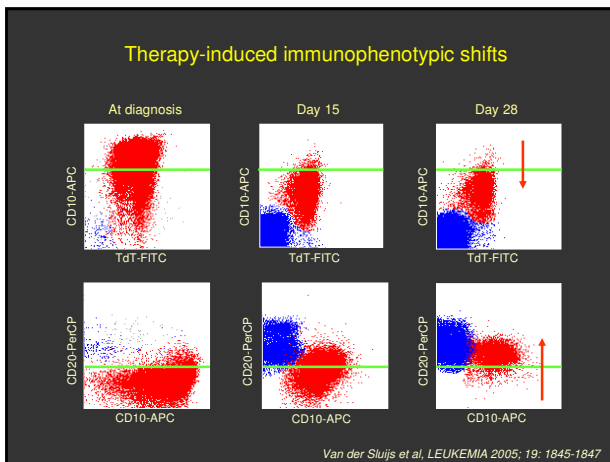
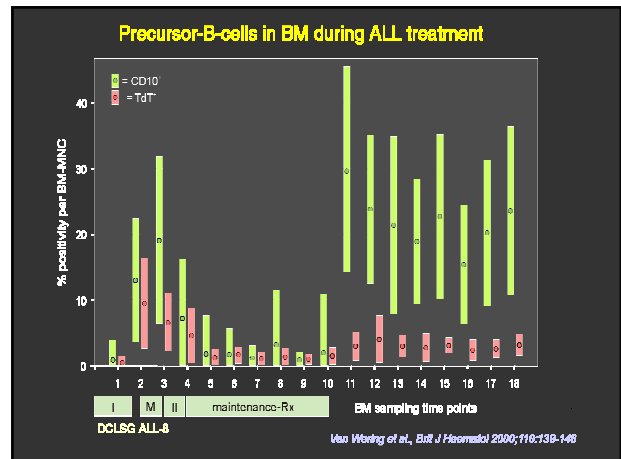
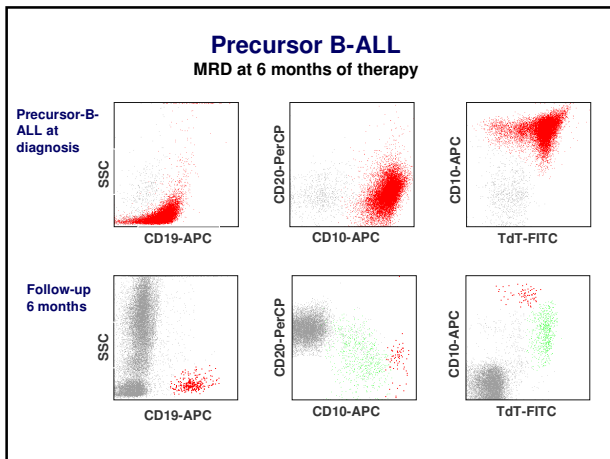
1. Quality Control Program: 2 times per year: - February / March
- August / September
2. Educational meetings including evaluation of quality control rounds: 2 times per year: - May / June
- October / November
3. Standardization of MRD techniques
 - standardization techniques within each treatment protocol
 - guidelines for interpretation of RQ-PCR results
4. Collaborative development and clinical evaluation of new MRD strategies and new MRD techniques

→ Basis for accreditation of MRD diagnostics




European Study Group on MRD detection Chairman: J.J.M. van Dongen





- ### Development of 8-color MRD panels
- Single-tube antibody EuroFlow MRD protocols under evaluation
- Acute leukemias (includes recognition of normal precursors)
 - Acute myeloid leukemia panel (AML-MRD): 1 tube per pathway (A. Orfao)
 - B-cell precursor (BCP-ALL-MRD): 1 tube (V. van der Velden, E. Mejstrikova)
 - T-cell ALL (T-ALL-MRD): 1 tube (V. Asnafi)
 - Chronic lymphoproliferative disorders (includes recognition of normal cells)
 - Chronic lymphocytic leukemia (CLL-MRD): 1 tube (A. Langerak)
 - Hairy cell leukemia (HCL-MRD): 1 tube (E. MacIntyre)
 - Mantle cell lymphoma (MCL-MRD): 1 tube (S. Böttcher)
 - Follicular lymphoma (FL-MRD): 1 tube (S. Böttcher)
 - Marginal zone lymphoma (MZL-MRD): 1 tube (P. Lucio)
 - Lymphoplasmacytic lymphoma (LPL-MRD): 1 tube (P. Lucio)
 - Diffuse large B-cell lymphoma (DLBCL-MRD): 1 tube (P. Lucio)
 - Burkitt lymphoma (BL): 1 tube (L. Lhermitte)
 - T-chronic lymphoproliferative diseases (T-CLPD-MRD): 1 tube (J. Almeida)
 - Multiple myeloma (MM): 1 tube (J. Flores)

Achievement of the EuroFlow Consortium: New concept in diagnostic flow cytometry



- Full technical standardization of multicolor flow cytometry (≥ 8 colors)
 - standardization of instrument settings and laboratory protocols
 - selection of fluorochromes and selection of antibody clones per marker
 - EuroFlow protocols work on all tested ≥ 8 colors flow cytometers:
 - DAKO Cyan, LSR-II, FACS Canto-II;
 - "late arrivals" (Navios and Gallios) still to be tested (new Workpackage)
- Implementation and further development of novel software: Infinicyt
 - fast and easy data handling with automated pattern recognition
 - combining multiple tubes: calculation and APS (principle component analysis)
 - mapping of diagnosis and follow-up leukemia samples against templates of "normal/control" samples
- Development of 8-color antibody protocols for diagnosis, classification and monitoring of hematological malignancies
 - 8-color panels are based on recognition of normal cells & differentiation pathways
 - diagnosis and classification tubes are ready; MRD tubes in development
 - flexibility within panels: deletion and inclusion of markers and tubes is possible
- Large EuroFlow data base linked to Infinicyt software

www.euroflow.org

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Photo Lukasz Sedek



MUCHAS GRACIAS